

# The principles of detection by sequencing

12:55-13:40

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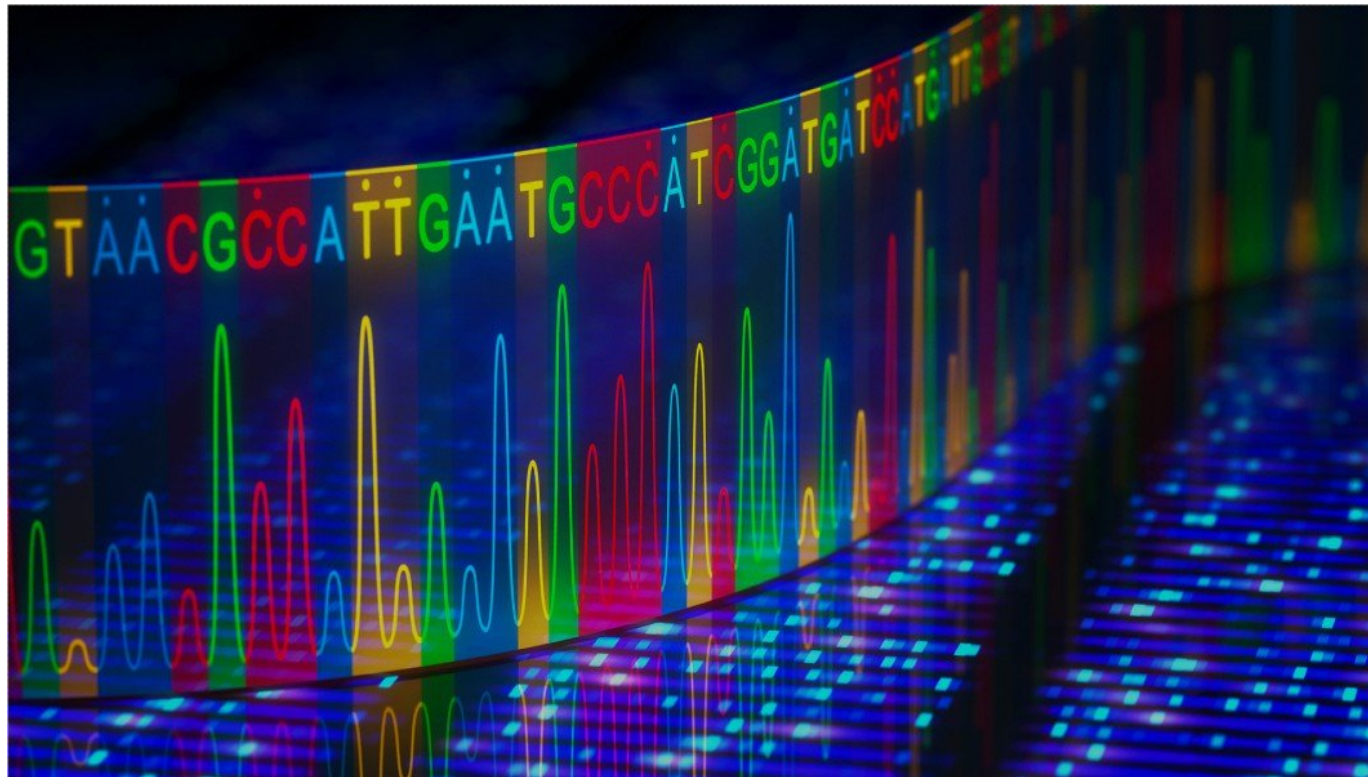
CTDNA PH.D. COURSE  
20 MAY 2021

AMANDA FRYDENDAHL BOLL JOHANSEN  
PHD STUDENT



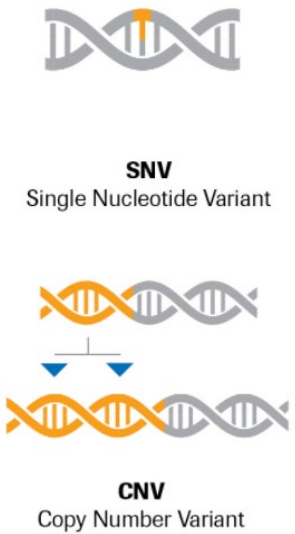
# NEXT GENERATION SEQUENCING

— Sequencing of DNA fragments in parallel

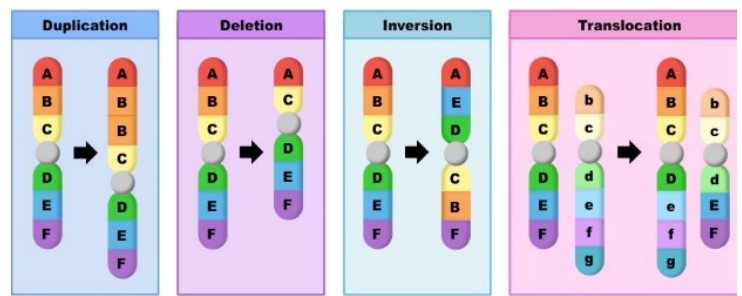


# TARGETS

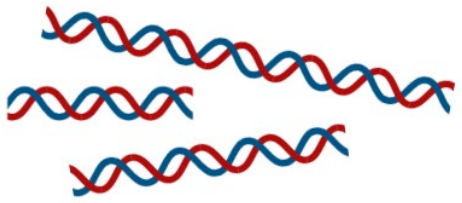
## Genetic variations



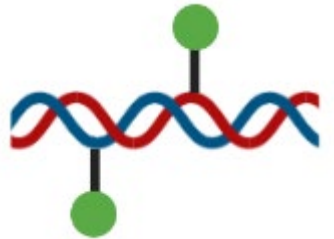
## Chromosomal changes



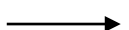
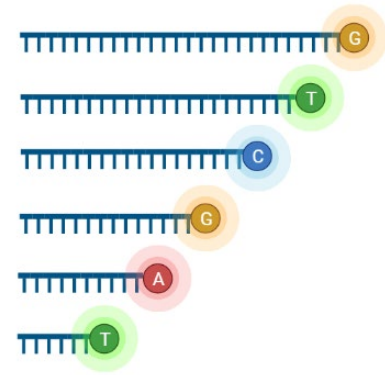
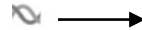
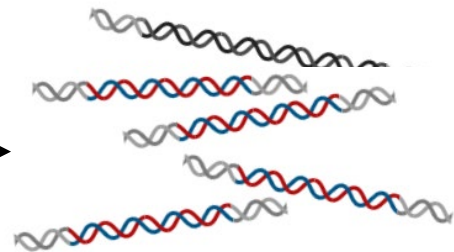
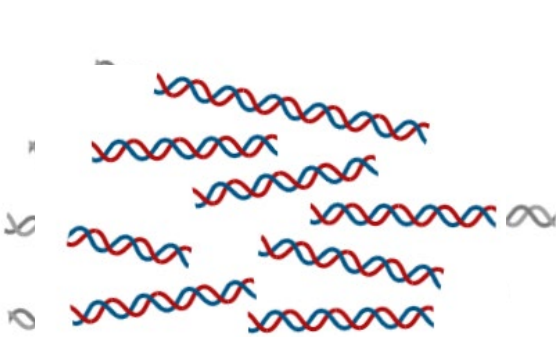
## Fragment length patterns



## DNA methylation



# NGS WORKFLOW



```
GATGGTCGTAGTCTTGA
      CGTAGTCTTGATGTCTGA
GATGGTCGTACTCTTGA
      GGTCGTAGTCTTGATGTCTGA
GAAGATGGACGTAGTCTTGA
.
.
.
```

## Library preparation, 1 day

- Addition of sequencing adapters to fragmented DNA
- PCR to amplify DNA

## (Capture), 2 days

- Focus analysis on regions of interest
- PCR to amplify DNA

## Sequencing, 1-2 days

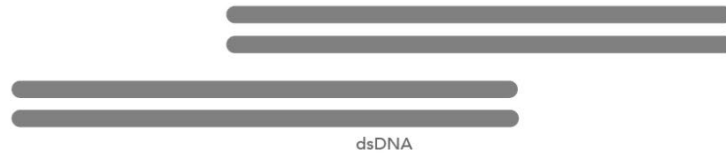
- Sequencing by synthesis
- Emits fluorescent signal

## Data analysis

- Align sequence data to reference genome
- Identify mutations

# LIBRARY PREPARATION

Fragmentation



End repair and A-tailing



Ligation



PCR amplification



Library



Adapter structure



P5 + P7

Binding to the sequencing flowcell

i5 + i7

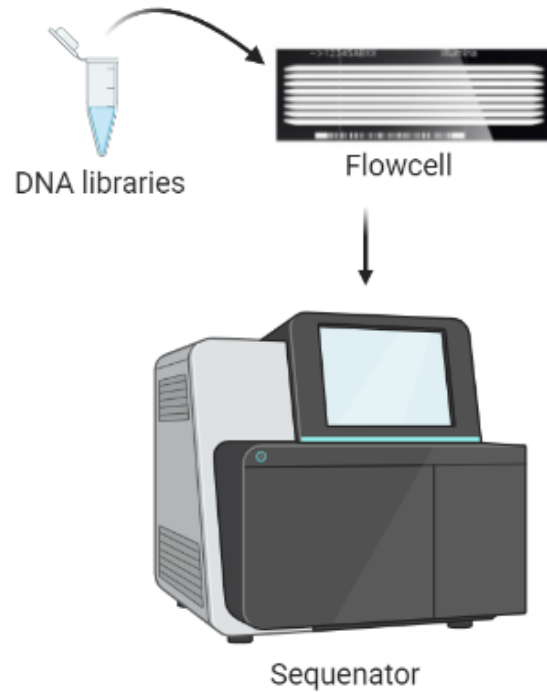
Sample-specific index sequences



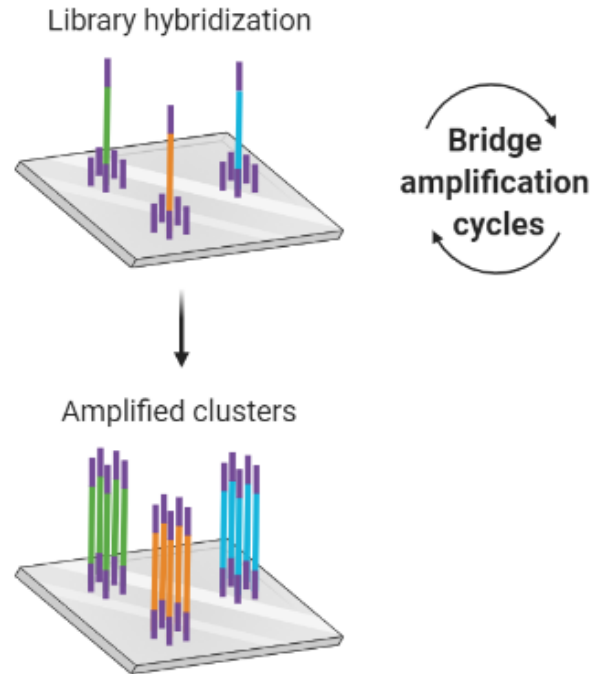
# ILLUMINA SEQUENCING

<https://www.youtube.com/watch?v=fCd6B5HRaZ8&t=19s>

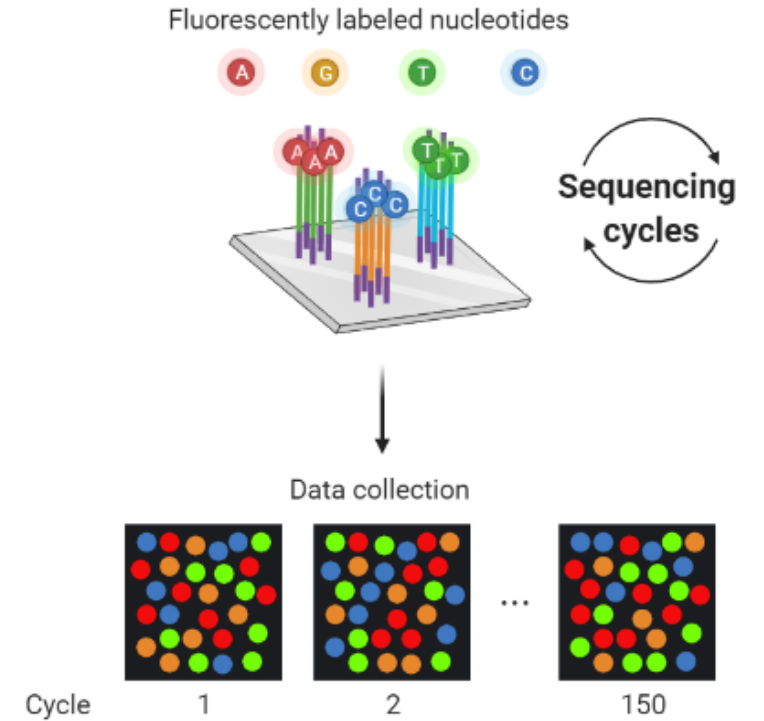
## ① DNA libraries loaded onto flowcell



## ② DNA library bridge amplification



## ③ DNA library sequencing



# (CAPTURE) Select regions of interest to focus analyses and minimize data output

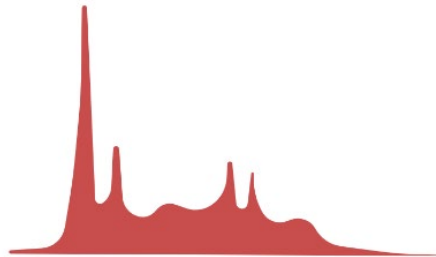
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## Whole genome sequencing (no capture)



Sequence entire genome

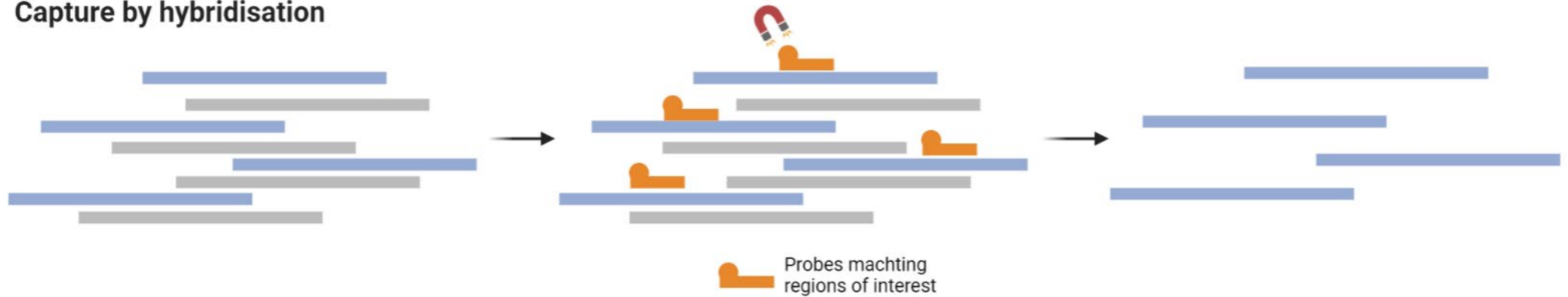
## Targeted sequencing (capture regions of interest)



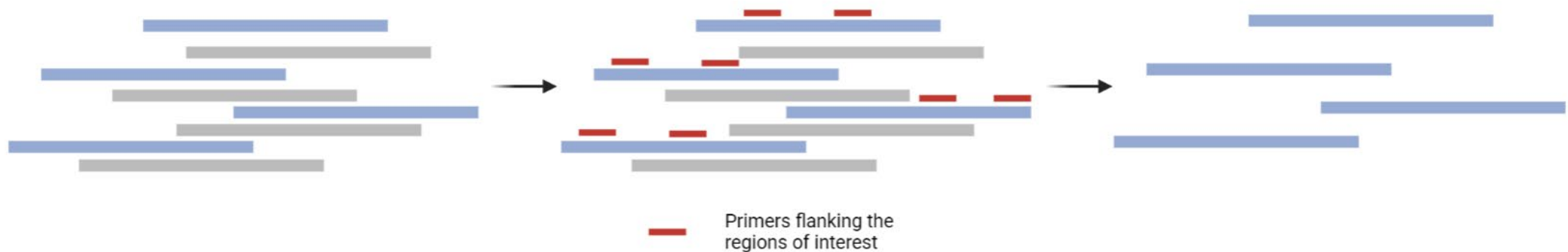
Focus analysis on regions of interest

# (CAPTURE) Select regions of interest to focus analyses and minimize data output

## Capture by hybridisation



## Capture by amplification





# (CAPTURE) Select regions of interest to focus analyses and minimize data output

Capture allow for higher sequencing depth at a fixed price

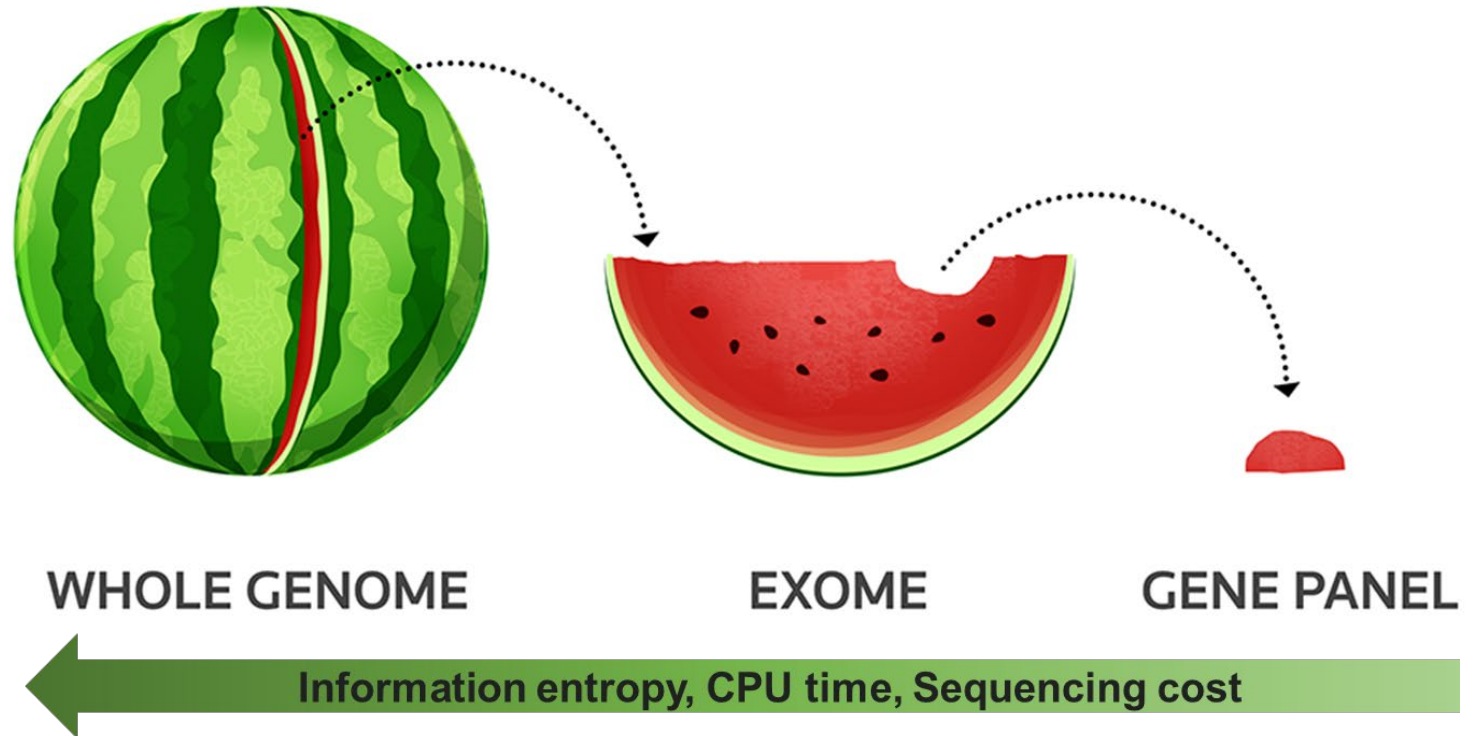
Whole genome sequencing at 1x:  
3.3 Gb (100% of genome)

or

Whole exome sequencing at 100x:  
33 Mb (1% of genome)

or

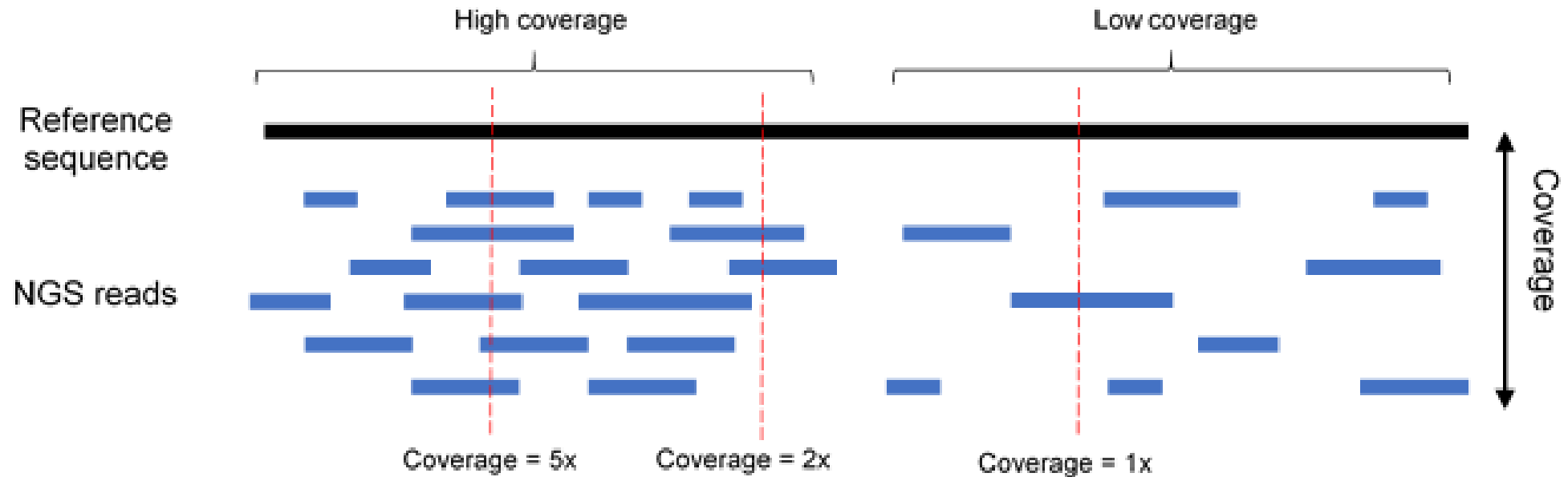
Targeted sequencing at 100,000x coverage :  
30,000 – 300,000 bp (0.001- 0.01% of genome)



# SEQUENCING COVERAGE

1000x coverage:

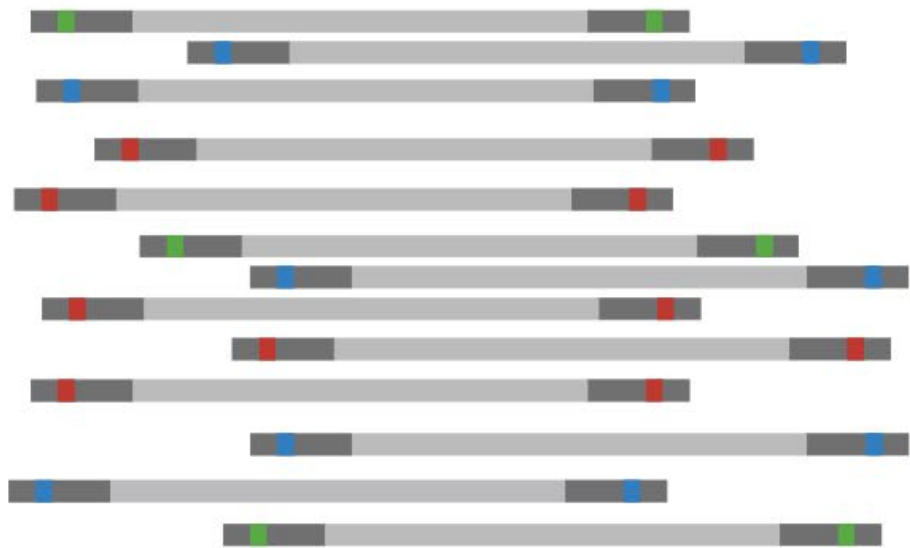
Possible to detect 1 in 1000 mutated reads



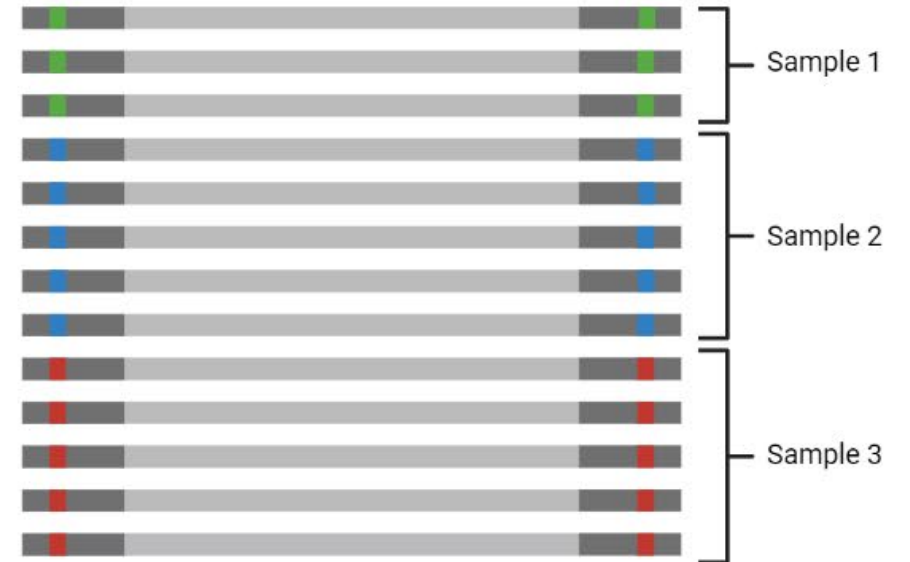
# MULTIPLEXING SAMPLES



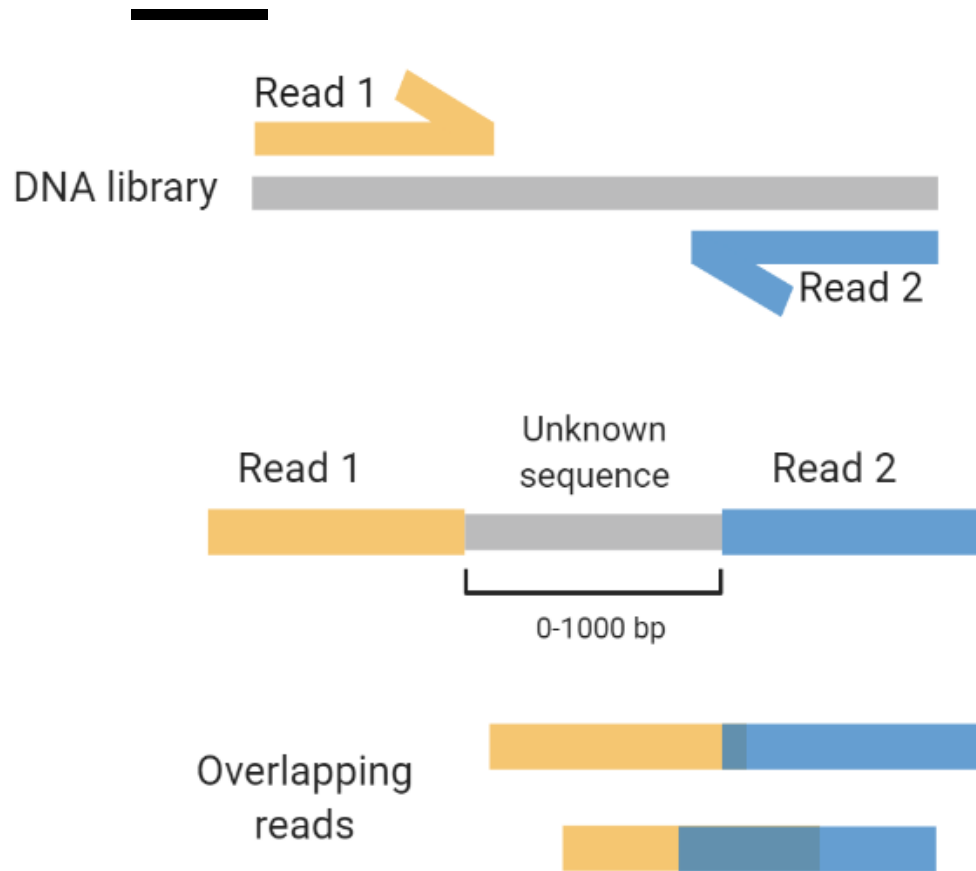
Adapter with  
sample-specific index sequence



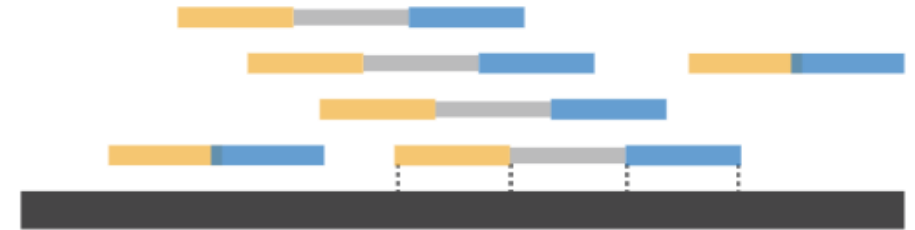
Demultiplexing accoring  
to index sequence



# PAIRED-END SEQUENCING



Reference sequence



- High quality alignment
- Determine fragment length

# ERRORS DURING SEQUENCING

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Errors can be introduced during:

- Samples storage
  - Sample processing, e.g. during PCR
  - Sequencing: PCR and optical errors
  - Data analysis: e.g. alignment
- } Action point: "unique molecular identifier" to each DNA fragment

→ Difficult to differentiate errors from true mutations → similar frequency

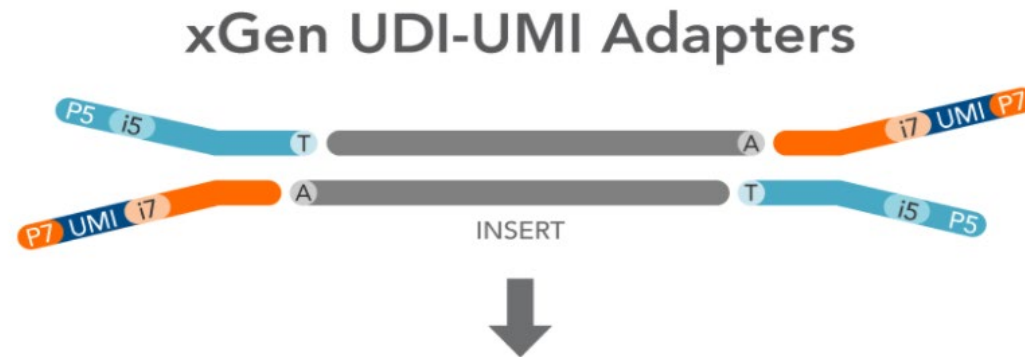
**RISK OF FALSE POSITIVE**

# UMI BASED ERROR CORRECTION

UMI = unique molecular identifier

- Random sequence of 3-9 nucleotides
- DNA molecule specific

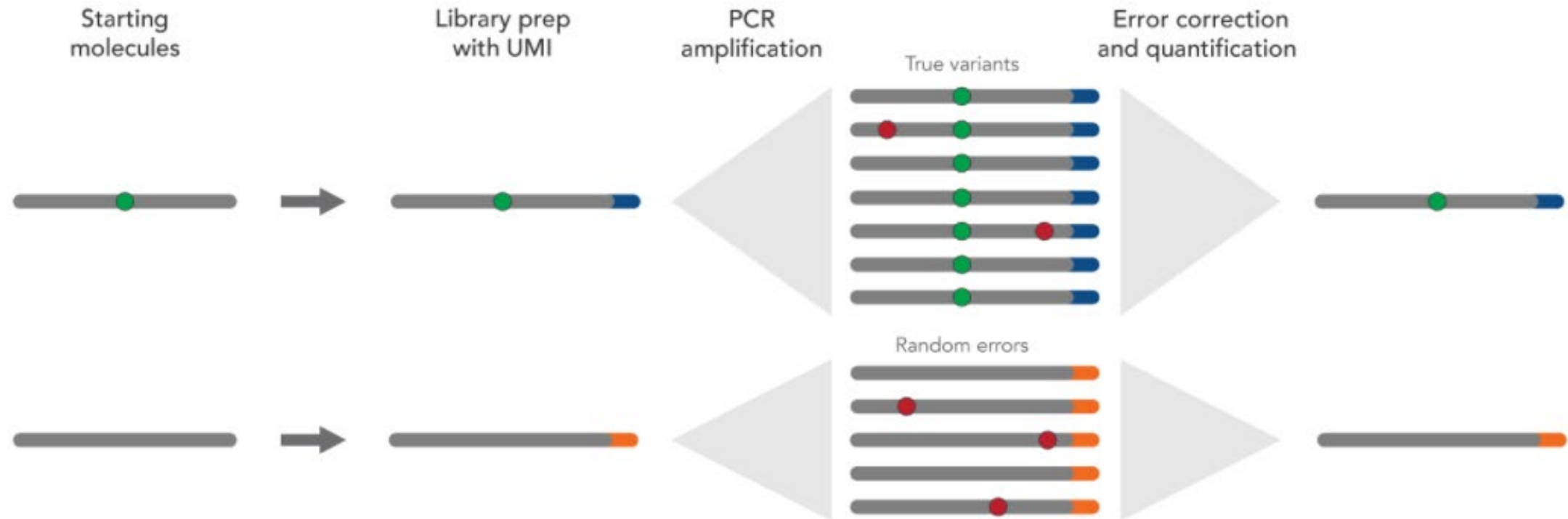
Ligation



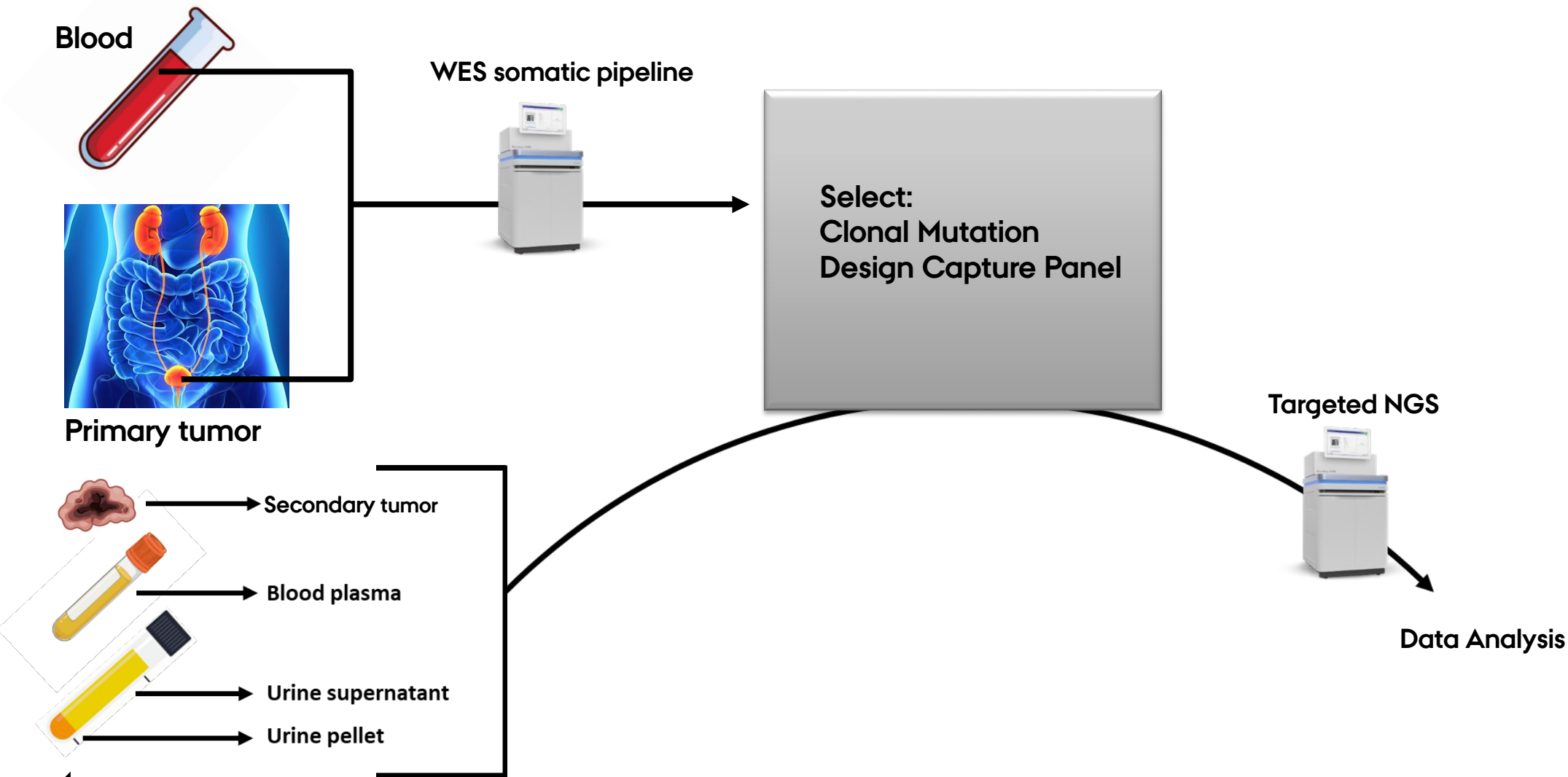
Library



# UMI BASED ERROR CORRECTION

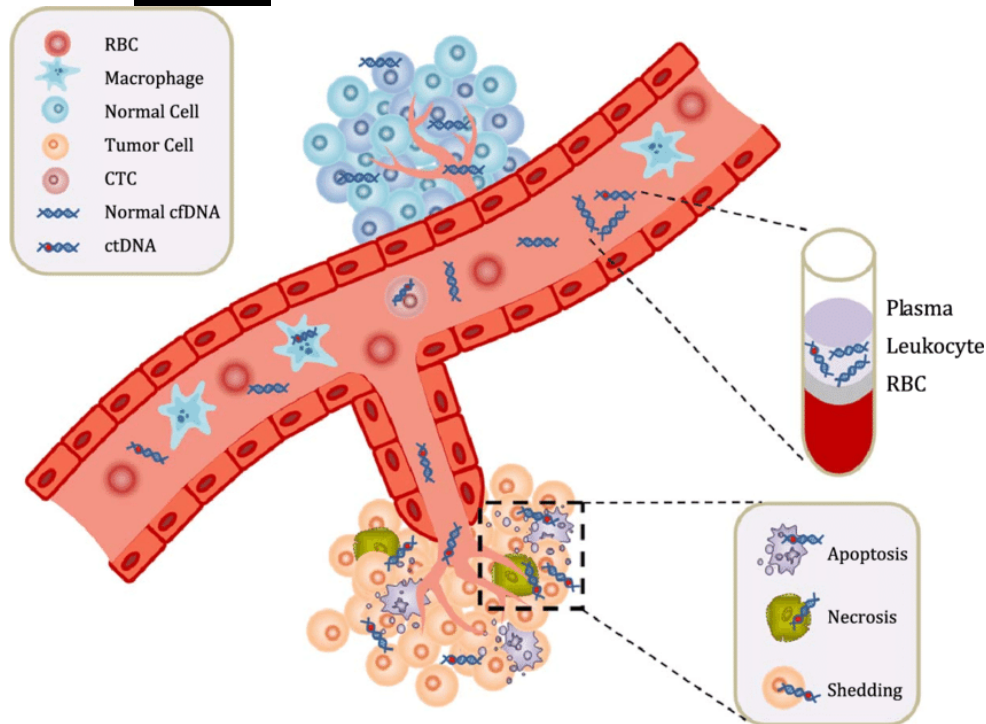


# WORKFLOW FOR TUMOR INFORMED PANEL SEQ FOR CANCER DISEASE MONITORING





# DETECT GENOMIC ALTERATIONS FROM CTDNA IN CFDNA FROM PLASMA



## Challenges

- Low amount of cfDNA (typically <50ng)
- The fraction of ctDNA in cfDNA are low.
- The frequency of tumor mutations in cfDNA can be very small
- Need ultra deep sequencing
- WGS and WES are costly

## Solution

- Tumor informed ddPCR assays
- Design small custom tumor informed panels
- Design small hotspot panels for specific cancers
- Include UMI to correct for increased PCR and NGS instrument bias

# WHEN TO SEQUENCE WHAT?

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## 1. cfDNA from liquid of choice (plasma, urine ect.)

- Self-explanatory: necessary for ctDNA detection
- Whole genome sequencing / targeted sequencing

## 2. DNA from normal cells (PBMC, normal tissue)

- Filter out patient-specific SNP's (VAF's around 50% or 100%, require coverage around 20-50x)
- Filter out CHIP mutations (VAF's around ~0.1%, require high coverage)

## 3. DNA from tumor

- Identify clonal targets for subsequent single-target analysis (e.g. ddPCR analysis)
- Generate a patient-specific mutational compendium to search for ctDNA



# GROUP DISCUSSION

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- Is sequencing useful in your projects for ctDNA detection?
- Can you think of applications where unique molecular identifies (UMIs) are not relevant/necessary?
- If you are/wish to use NGS in your projects, is sequencing of the tumor and/or normal cells necessary? Why? Why not?





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